

FluxOR™ Potassium Ion Channel Assay

Catalog Number F10016, F10017

Doc. Part No. 100007420 Pub. no. MAN0002485 Rev. A.0

Detailed protocol is available online at www.thermofisher.com.

Protocol summary

1. For each microplate, prepare 10 mL of **Loading Buffer**:

PowerLoad™ Concentrate	100 µL
FluxOR™ Reagent, reconstituted in DMSO	10 µL
Deionized water	8.8 mL
FluxOR™ Assay Buffer	1 mL
Probenecid, reconstituted in deionized water	100 µL
Total volume	10 mL

2. Remove media from cells and add 20 µL (for 384-well plate) or 80 µL (for 96-well plate) of **Loading Buffer** to each well.

3. Incubate for 60 minutes at 18–24°C, protected from direct light.
During incubation, prepare the **Assay** and **Stimulus Buffers**.

4. Prepare 10 mL of **Assay Buffer**:

Deionized water	8.9 mL
FluxOR™ Assay Buffer	1 mL
Probenecid, reconstituted in deionized water	100 µL
Total volume	10 mL

Protocol continued on reverse side

5. Prepare 5 mL of **Stimulus Buffer**:*

	+K ⁺	-K ⁺
Deionized water	2.5 mL	3.5 mL
FluxOR™ Chloride-free Buffer	1 mL	1 mL
K ₂ SO ₄ Concentrate	1 mL	—
Tl ₂ SO ₄ Concentrate	0.5 mL	0.5 mL
Total volume	5 mL	5 mL

* Additional Stimulus Buffer may be required based on liquid handling capabilities.

6. Remove **Loading Buffer** and replace with 20 µL per well (for 384-well plate) or 80 µL per well (for 96-well plate) of **Assay Buffer**.

7. *Optional*: Add the test compounds, then incubate for 10–30 minutes at 18–24°C.

8. Perform assay using a fluorescent plate reader with liquid handling features.

- Use a FITC filter or set the excitation/emission wavelengths to ~490 nm/~525 nm.
- Add **Stimulus Buffer** after 10 seconds of recording (5 µL for 384-well plate or 20 µL for 96-well plate).
- Read plate every 1–2 seconds for 1–3 minutes.

For more information, visit www.thermofisher.com/fluxor.

The information in this guide is subject to change without notice.

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