

GC-MS/MS Analysis of Benzodiazepines Using Analyte Protectants

Jeremy Matthews,¹ Alex Chen,² and Flavio Bedini¹

¹Thermo Fisher Scientific, Singapore; ²Alpha Analytical Pte Ltd, Singapore



Overview

Purpose: Assess the feasibility of using analyte protectants to improve the analysis of benzodiazepines.

Methods: Benzodiazepines were analyzed using triple-quadrupole GC-MS/MS. The mass spectrometer was run in EI mode using 3 SRM transitions per analyte.

Results: The calibration linearity of the benzodiazepines varied significantly between compounds; diazepam showed excellent linearity while lorazepam, lectopam, clonazepam and nitrazepam showed very poor responses. Sorbitol was used as analyte protectant to significantly improve the calibration behavior of the more challenging analytes.

Introduction

Benzodiazepines drugs are psychoactives whose structure contains a benzene ring fused with a diazepine ring. One of the most well known benzodiazepines – diazepam – has been marketed under the name “Valium” since 1963. These drugs are effective tranquilizers and as such are commonly used in medication to treat anxiety and sleep disorders amongst other conditions. The relative availability of these drugs combined with their sedative effect has led to their illicit use as either recreational drug and sometimes in suicides. Consequently, it is common practice to analyze for benzodiazepines in forensic and toxicology laboratories.

Benzodiazepines compounds are bases and readily react with active sites in eg. the GC inlet liner causing problems in analysis at low levels and resulting in poor linearity and reproducibility. The use of analyte protectants reduces liner activity and often enables the detection of such ‘active’ compounds at much lower levels. The use of analyte protectants in the GC-MS analysis of a group of benzodiazepines was investigated and the results are discussed below.

Methods

Analytes

The following benzodiazepines were analyzed in this work: lorazepam, diazepam, lectopam, nitrazepam, clonazepam. Standards of these analytes were spiked into a matrix.

GC-MS System

All measurements were carried out using the Thermo Scientific™ TSQ™ 8000 triple quadrupole GC-MS/MS system equipped with the Thermo Scientific™ TRACE™ 1310 GC with SSL Instant Connect SSL module and Thermo Scientific™ TriPlus™ RSH autosampler. The method details are given in Table 1 below.

TABLE 1. Instrument method for benzodiazepine analysis

TRACE 1310 GC	
Injection mode	splitless
Splitless Time	1.0 min
GC Column	Trace Gold TG-5SilMS, 15 m × 0.25 mm × 0.25 µm
Carrier gas	He (99.999 %)
Flow	1.2 mL/min, constant flow
Temperature program	50 °C, 2 min 20 °C/min to 300 °C, 2 min
Transfer line temperature	280 °C
Total analysis time	14.6 min
TriPlus RSH Autosampler	
Injection volume	1 µL
TSQ 8000 MS/MS	
Ionization mode	EI, 70 eV
Ion source temperature	250 °C
Scan mode	SRM using timed SRM
SRM transition setup	automatically build-up by AutoSRM software, transitions see Table 2

Results

AutoSRM study

Benzodiazepine standards were purchased, and calibration solutions were prepared between 10 and 10000 ppb. The 100 ppb solution was then used for an AutoSRM study to determine the optimal SRM transitions and for each compound, and thereby create the MS/MS method.

AutoSRM is a unique MS/MS method development tool included in the TSQ 8000 GC-MS/MS software suite which uses a vial containing a standard solution of the compounds required for the method, in the TriPlus RSH autosampler and automatically optimizes retention time, precursor and product masses and collision energy for quant and confirming ions.

The AutoSRM study was completed with only minimal user interaction required, and the optimized SRM transitions are shown in Table 2 below.

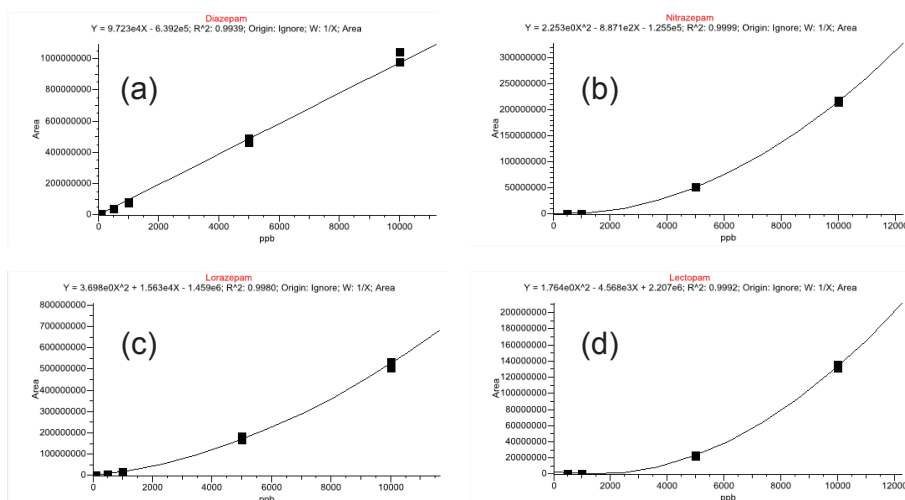
TABLE 2. Instrument method for benzodiazepine analysis

Compound	CAS Number	RT (min)	Precursor Mass (m/z)	Product Mass (m/z)	Collision Energy (eV)
Lorazepam	846-49-1	14.32	239.0	176.7	25
Lorazepam	846-49-1	14.32	274.0	239.1	10
Diazepam	439-14-5	14.45	256.1	165.1	30
Diazepam	439-14-5	14.45	256.1	221.1	10
Lectopam	1812-30-2	15.32	316.7	288.0	20
Lectopam	1812-30-2	15.32	316.7	289.1	10
Nitrazepam	146-22-5	16.02	280.3	205.1	30
Nitrazepam	146-22-5	16.02	280.3	234.1	10
Clonazepam	1622-61-3	16.51	280.0	234.1	10
Clonazepam	1622-61-3	16.51	314.0	268.1	15

Benzodiazepine Calibration Curves

The optimized MRM method created using AutoSRM (described above) was used to record calibration data for the benzodiazepines at up to 10 000 ppb. The linearity of the calibration curves varied widely with the particular analyte as shown in Figures 1 (a) – (d) below.

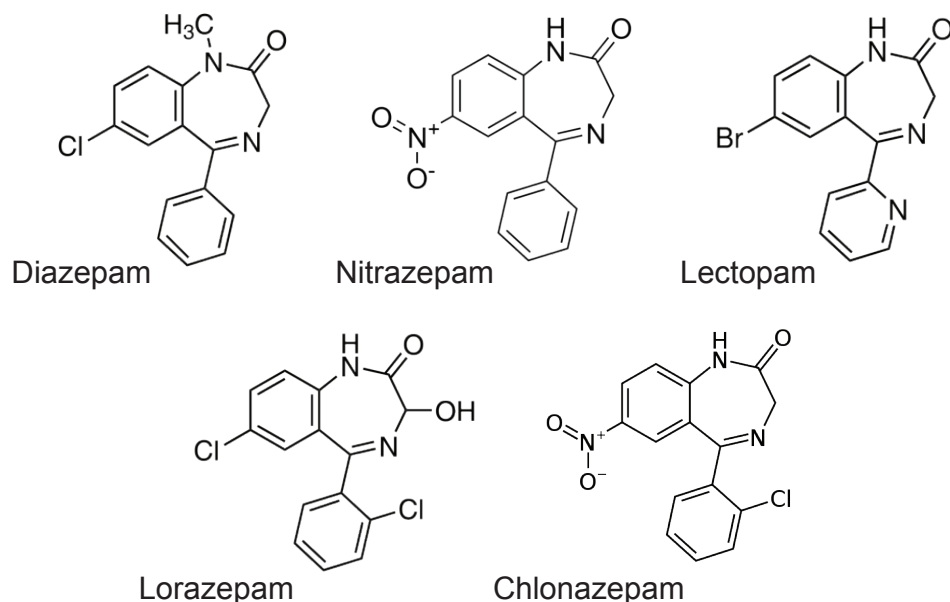
FIGURE 1. Calibration curves for (a) diazepam, (b) nitrazepam, (c) lorazepam and (d) lectopam at up to 10,000 ppb.



The calibration curves for diazepam was very linear ($R^2 > 0.99$) between 5 and 10,000 ppb also exhibited excellent peak shape. The other benzodiazepines, however, strongly deviated from linearity, and a quadratic was required to achieve a reasonable fit of the data (also for clonazepam, not shown). The peak areas for these compounds were also fairly low and showed poor signal-to-noise at relatively high concentrations.

The difficulties encountered in analysis of such polar compounds have been previously reported in the analysis of pesticides^{1, 2, 3}. The high 'activity' of polar functional groups has been observed to lead to poor recoveries and significant loss of analytes at low concentration due to trapping and degradation at active sites in the gas flow path. The degradation of trapped compounds from previous injections can then form further active sites (increased surface area), which may rapidly lead to poor chromatographic response and performance.

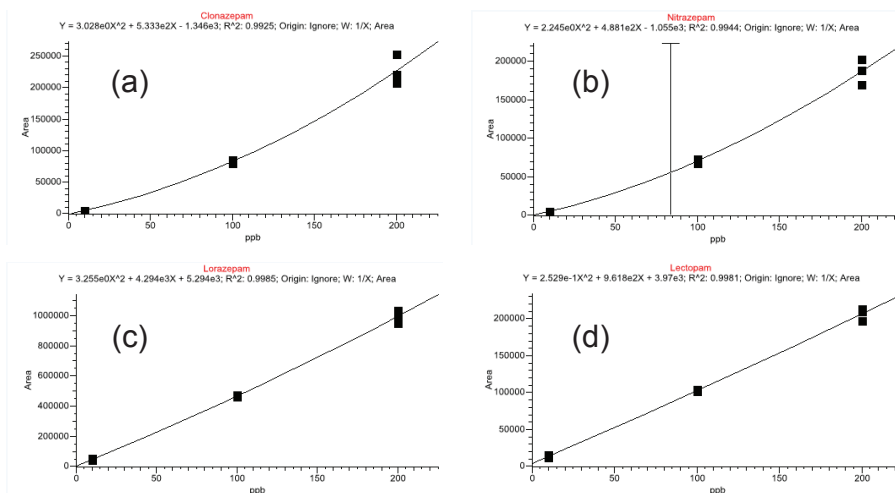
FIGURE 2. Chemical structure of the benzodiazepine compounds analyzed in this work.



It has been previously shown that the use of analyte protectants, such as sorbitol is very effective in correcting this behavior for eg. pesticide analysis, whereby the protectant reacts with active sites in the GC flowpath effectively deactivating the system and enhancing analyte recovery, especially at low levels.

This has several advantages including improving peak shape (less tailing) and hence repeatability (RSDs), as well as keeping the inlet liner clean for longer durations as analyte and matrix are no longer trapped and subsequently degraded.

FIGURE 3. Calibration curves for (a) clonazepam, (b) nitrazepam, (c) lorazepam and (d) lectopam between 10 and 200 ppb using 0.2 % sorbitol as analyte protectant.



Analyte protectant (0.2% sorbitol) was added to the benzodiazepine calibration standards, and the calibration curves were measured as shown above in Figure 3. The calibration linearity improved dramatically for lorazepam, lectopam, clonazepam and nitrazepam with the sorbitol addition. The response factors for lectopam and lorazepam increased by roughly 16 and 7 times respectively indicating a dramatic improvement in sensitivity.

Conclusions

- Benzodiazepines are challenging to analyze by GC-MS/MS due to high activity as a result of polar functional groups.
- Activity of inlet liner and GC column result in adsorption/degradation of some benzodiazepine analytes leading poor linearity of calibration and sensitivity.
- The use of sorbitol as analyte protectant dramatically improved the linearity of the response, as well as signal intensity for these compounds.
- All benzodiazepines could be quantified at 10 ppb using analyte protectant.

References

1. Anastassiades, M., Maštovská, K., Lehotay, S.J., Evaluation of analyte protectants to improve gas chromatographic analysis of pesticides, *J. Chromatogr. A* 1015 (2003) 163-184.
2. Mastovska, K., Lehotay, S.J., Anastassiades, M., Combination of Analyte Protectants To Overcome Matrix Effects in Routine GC Analysis of Pesticide Residues in Food Matrixes, *Anal. Chem.* 77(24) (2005) 8129-8137.
3. Li, Y., Chen, X., Fan, C., Pang, G., Compensation for matrix effects in the gas chromatography–mass spectrometry analysis of 186 pesticides in tea matrices using analyte protectants, *J. Chrom. A* 1266 (2012), 131-142.

www.thermoscientific.com

©2014 Thermo Fisher Scientific Inc. All rights reserved. ISO is a trademark of the International Standards Organization. All other trademarks are the property of Thermo Fisher Scientific Inc. and its subsidiaries. This information is presented as an example of the capabilities of Thermo Fisher Scientific products. It is not intended to encourage use of these products in any manners that might infringe the intellectual property rights of others. Specifications, terms and pricing are subject to change. Not all products are available in all countries. Please consult your local sales representative for details.



Thermo Fisher Scientific,
Austin, TX USA is
ISO 9001:2008 Certified.

Africa +43 1 333 50 34 0
Australia +61 3 9757 4300
Austria +43 810 282 206
Belgium +32 53 73 42 41
Canada +1 800 530 8447
China 800 810 5118 (free call domestic)
400 650 5118

Denmark +45 70 23 62 60
Europe-Other +43 1 333 50 34 0
Finland +358 9 3291 0200
France +33 1 60 92 48 00
Germany +49 6103 408 1014
India +91 22 6742 9494
Italy +39 02 950 591

Japan +81 45 453 9100
Latin America +1 561 688 8700
Middle East +43 1 333 50 34 0
Netherlands +31 76 579 55 55
New Zealand +64 9 980 6700
Norway +46 8 556 468 00
Russia/CIS +43 1 333 50 34 0

Singapore +65 6289 1190
Spain +34 914 845 965
Sweden +46 8 556 468 00
Switzerland +41 61 716 77 00
UK +44 1442 233555
USA +1 800 532 4752

Thermo
SCIENTIFIC

A Thermo Fisher Scientific Brand