

Influence of Gender and Ethnicity on Drug Metabolizing Enzymes in Primary Human Hepatocytes



AB applied biosystems | invitrogen

Matthew Sherman, Cornelia Smith, Heather Rollins, Jeanine Fogarty and Jasminder Sahi

Cell Systems Division • Life Technologies • 4301 Emperor Blvd. • Durham, North Carolina 27560 • USA

Introduction

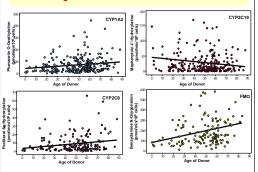
Marked differences are observed in pharmacokinetics of drugs between individuals, resulting in decreased efficacy or adverse drug reactions. Defective alleles encoding biotransformation enzymes such as CYP450s and UGTs can result in decreased drug clearance, thereby increasing the incidence of toxicity with many drugs. In some cases, polymorphic drug metabolizing enzymes may enhance efficacy e.g. omeprazole levels are maintained more efficiently in patients carrying certain alleles for CYP2C19¹. Conversely, variant alleles of UGT1A1 are less capable of conjugating and eliminating SN-38, the active form of irinotecan, potentially causing toxicities². Understanding the drug metabolizing enzymes responsible for metabolism and more importantly, elucidating the genetic basis for this variation would make drug therapy safer and more efficacious.

Polymorphism are reported in CYP2D6, 2C19, and 2C9, 1A1, 2A6, 2A13, 2C8, 3A4, and 3A5. Approximately 5-14% of Caucasians, 0-5% Africans, and 0-1% of Asians are poor CYP2D6 metabolizers. CYP1A1/2 play a role in procarcinogen bioactivation and polymorphisms can contribute to the variable susceptibility to carcinogenesis. People of African descent require higher tacrolimus doses than Caucasians to reach similar trough concentrations of Tacrolimus³.

Sex-specific differences have also been observed where women are reported to experience more adverse drug effects than men e.g. women are more likely to experience drug-induced QTprolongation and torsade-de-pointes arrhythmia4. Saguinavir and indinavir treatment results in clinically significant sex differences in plasma drug concentrations and clinical outcomes, with women having higher drug exposure4. Pharmacokinetics of antidepressant drugs can be substantially different, with women responding better to MAOI antidepressants and men responding better to TCAs⁵. Clinical studies indicate that CYP2E1 and CYP1A2 activity is higher in males, while CYP3A appears to have greater activity in females⁶. However, this is difficult to confirm in vivo, as multiple factors affect drug metabolism.

The molecular basis for many of these sex- and ethnicity- based differences have yet to be elucidated. In a move towards understanding these, we have characterized 254 preparations of primary human hepatocytes for the major wild type and polymorphic drug metabolizing enzymes and evaluated the influence of ethnicity, gender, age and BMI on activities.

Figure 1 - The Effect of Age on **Drug Metabolizing Enzyme** Activity



CYP1A2: Increase in activity with increasing donor age. p = 0.004; r = 0.18; n = 249

CYP2C19: Decrease in activity with increasing donor age. p = 0.075: r = -0.11: n = 245CYP2C8: Increase in activity with increasing donor age.

p = 0.008: r = 0.201: n = 175FMO: Increase in activity with increasing age. p =

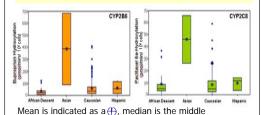
0.001; r = 0.281; n = 133Phase I enzymes: CYP2D6, CYP2B6, CYP2C9, CYP2E1, and CYP3A did not show any statistically significant changes with age of donor.

Phase II enzymes: SULT and UGT did not show statistically significant changes with age of donor.

Statistics determined by linear regression (Pearson

The correlation between drug metabolizing enzyme activity and age is plausibly non-zero.

Figure 2 – The Effect of **Ethnicity on Drug Metabolizing Enzyme Activity**



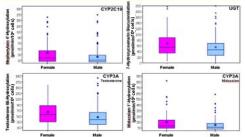
horizontal line in the box, outliers are indicated with an asterisk. Statistics determined by ANOVA. CYP2B6: Higher activity in Asian vs. all other ethnicities studied, followed by Hispanic > Caucasian > African descent. p = 0.00; n = 15 African descent, 2 Asian, 141 Caucasian, 4 Hispanio

CYP2C8: Higher activity in Asian > Hispanic > African descent > Caucasian. p = 0.00; n = 15 African descent, 2 Asian, 155 Caucasian, 3 Hispanic UGT activity different between Caucasian and African

CYP1A2, CYP2C9, CYP2C19, CYP2D6, CYP2E1, CYP3A, SULT, FMO: No marked differences in activities between these different ethnicities.

Ethnicity has a non-zero effect on certain drug metabolizing enzyme activities in primary human hepatocytes.

Figure 3 - The Effect of Gender on Drug Metabolizing Enzyme Activity



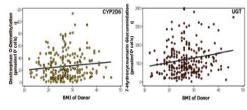
Mean is indicated as a \oplus , median is the middle horizontal line in the box, outliers are indicated with an asterisk. Statistics determined by ANOVA. CYP2C19: Higher activity in female vs. male primary hepatocytes. p = 0.06: n = 118 female and 127 male UGT: Higher activity in female vs. male primary hepatocytes p = 0.03; n = 116 female and 138 male CYP3A4 (Testosterone): Higher activity in female vs. male primary human hepatocytes. p = 0.002; n = 119female and 130 male

CYP3A4 (Midazolam): Higher activity in female vs. male primary human hepatocytes. p = 0.05; n = 89 female

CYP1A2, CYP2B6, CYP2C8, CYP2C9, CYP2D6, CYP2E1, SULT, FMO: No major changes in Phase I or Phase II drug metabolizing activity by gender

The correlation between drug metabolizing enzyme activity and gender is plausibly non-zero.

Figure 4 – The Effect of Body Mass Index (BMI) on Drug **Metabolizing Enzyme Activity**

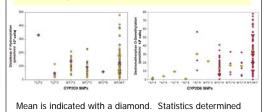


CYP2D6: Trend towards increase in activity with increasing BML p = 0.07: r = 0.114: p = 248UGT: Increase in activity with increasing age. p = 0.014: r = 0.154: n=254CYP1A2, CYP2B6, CYP2C19, CYP2C8, CYP2C9, CYP2E1, CYP3A, SULT, FMO: No marked changes in Phase I or

Phase II drug metabolizing activity at a range of BMI. Statistics determined by linear regression (Pearson

BMI does not appear to affect activity of the major drug metabolizing enzymes.

Figure 5 – The Effect of SNPs on **Drug Metabolizing Enzyme Activity**



by ANOVA.

There are statistical differences in CYP2C9 activity between hepatocytes with certain CYP2C9 SNPs. In CYP2C9, *2/*3, WT/*3, and WT/*6 have lower activity than the wild type. *2/*2 and WT/*2 have higher activity than the wild type (p = 0.06). CYP2C9: n = 1 *2/*2, 19 WT/*2, 5 *2/*3, 17 WT/*2, 1WT/*6, 93 WT/WT

Details of Donor with CYP2C9 *2/*2 polymorphism

Statistically lower CYP2D6 activity was observed in hepatocytes that had SNPS for CYP2D6. p = 0.00Presence of any CYP2D6 polymorphism decreases CYP2D6 activity.

CYP2D6: n = 7 * 4/* 4, 2 * 6/* 6, 3 * 9/* 9, 1 WT/* 3, 42WT/*4, 1 *4/*6, 1 *4/*9, 4 WT/*6, 6 WT/*9, 94

The presence of SNPs has a non-zero effect on drug metabolizing enzyme activity

Figure 6 - Cryopreserved Hepatocytes





Table 1 - Case Study of a Single **Donor**

26 year old Caucasian Female SNPs: CYP2C9*2/*2; CYP3A5 *3/*3

CYP2C9*2/*2 activity: 332 pmoles/min/mg protein (mean of 128 in wt/wt) CYP3A5 "3/"3: 1700 pmoles/min/mg protein (mean of 236 in wt/wt, mean of 95.9 in all other donors with "3/"3) ECOD: 31.9 pmoles/min/mg/protein (mean of 34.9 in population)

BMI: 26 Alcohol: social

Smoker: 1 packet / day for 10 years

Prescriptions for 5 years: Effexor, Lortab, Oxycontin, Clonidine

Materials and Methods

Hepatocyte Isolation and Cryopreservation. Human hepatic tissues were derived from transplant reject livers and liver resections from healthy tissue (sourced through the Hepatic Research Registry), mainly from patients with hepatic cancer. Primary human hepatocytes were isolated from liver tissue by a two-step collagenase perfusion method⁷ and subsequently cryopreserved. Cryopreserved hepatocytes were thawed in serum-containing medium, re-suspended in serum-free incubation medium, and assessed for cell viability by Trypan blue exclusion.

Metabolic Activity. Enzyme activities were determined in suspension hepatocytes at 0.5 x106 cells/mL in WEM with the probe substrates: phenactin (CYP1A2), bupropion (CYP2B6), paclitaxel (CYP2C8), diclofenac (CYP2C9), S-mephenytoin (CYP2C19), dextromethorphan (CYP2D6), chlorzoxazone (CYP2E1), testosterone & midazolam (CYP3A), benzydamine (FMO), 7-ethoxycoumarin (ECOD) and 7-hydroxycoumarin (UGT & SULT). Metabolites were identified by LC/MS/MS analysis.

Genotyping. Genotyping for thirteen different single nucleotide polymorphisms (SNPs) within four different genes, CYP2C9, CYP2C19, CYP2D6, CYP3A5, were detected by DNA isolation and ABI TagMan® primer/probe sets.

Data Analysis. ANOVA and Pearson correlations (linear regression), were used to determine relationships between variables. The level of significance was 0.05 ($\alpha = 0.05$). The variables were age, gender, ethnicity, BMI, presence of polymorphisms, Phase I drug metabolizing enzymatic activity (listed above), and Phase II drug metabolizing enzymatic activity (listed above).

Conclusions

- •272 donor livers were evaluated for SNPs and primary hepatocyte activities
- •Plausibly non-zero relationships between age and metabolic activities of CYP1A2 (increase), CYP2C8 (increase), FMO (increase); moderate decrease of CYP2C19, and no effect on CYP2D6, CYP2B6, CYP2C9, CYP2E1, CYP3A, SULT and UGT as determined by Pearson correlation.
- •Minimal effect of BMI on drug metabolizing enzyme activity - moderate increase in CYP2D6 and a marked increase in UGT activities as determined by Pearson
- •Gender affects activity of some of the major drug metabolizing enzymes, with higher activities observed in females for UGT and CYP3A4, moderately higher activities in CYP2C19 and no differences in CYP1A2. CYP2B6, CYP2C8, CYP2C9, CYP2D6, CYP2E1, SULT & FMO as determined by ANOVA.
- •Ethnicity of donor liver has an effect on enzymes in hepatocyte preparations as determined by ANOVA:
 - •CYP2B6 activity: Asian >> Hispanic > Caucasian > African descent
 - •CYP2C8 activity: Asian >> Hispanic > African descent > Caucasian
 - •UGT activity: Caucasian >> African descent (p = 0.03).
- •Statistically different enzyme activity observed in hepatocytes from livers with certain SNPs as determined by ANOVA. Drug use could change this profile.
- © 2010 Life Technologies Corporation. All rights reserved.

The trademarks mentioned herein are the property of Life Technologies Corporation or their respective owners. TagMan is a registered trademark of Roche Molecular

For Research Use Only. Not intended for any animal or human therapeutic or diagnostic use.

References

- (1) Crettol, S.; Petrovic, N.; Murray, M. Curr Pharm Des, 16, 204-19.
- (2) Cecchin, E.; Innocenti, F.; D'Andrea, M.; Corona, G.; De Mattia, E.; Biason, P.; Buonadonna, A.; Toffoli, G. J Clin Oncol 2009, 27, 2457-65
- (3) Yasuda, S. U.; Zhang, L.; Huang, S. M. Clin Pharmacol Ther 2008, 84, 417-23.
- (4) Thurmann, P. A. Ther Umsch 2007, 64, 325-9.
- (5) Bies, R. R.; Bigos, K. L.; Pollock, B. G. J Gend Specif Med 2003, 6, 12-20.
- (6) Scandlyn, M. J.; Stuart, E. C.; Rosengren, R. J. Expert Opin Drug Metab Toxicol 2008, 4, 413-24.
- (7) LeCluyse, E. L.; Alexandre, E.; Hamilton, G. A.; Viollon-Abadie, C.; Coon, D. J.; Jolley, S.; Richert, L. Methods Mol Biol 2005, 290, 207-29.