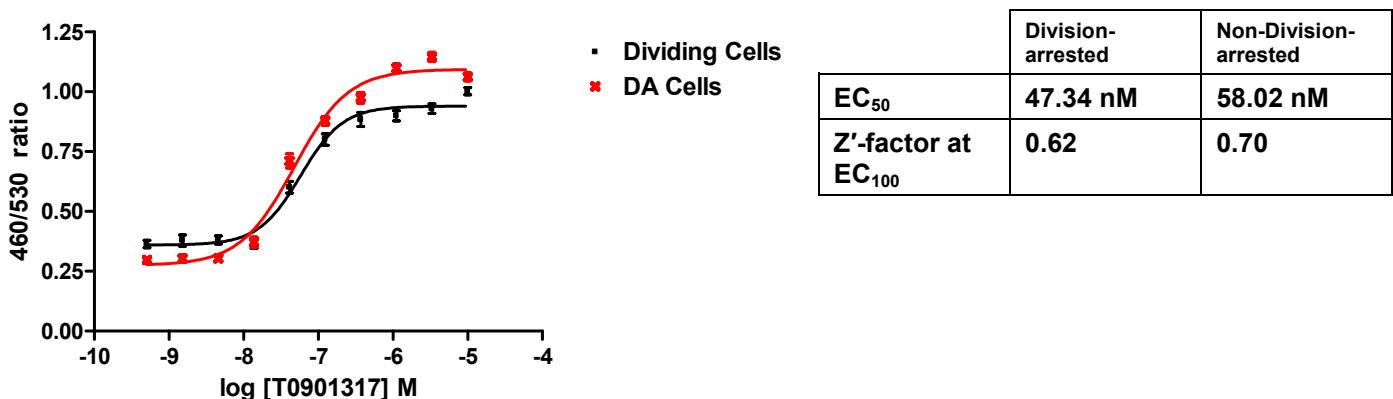


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## 1. Description

GeneBLazer® LXR beta HEK 293T DA (Division-arrested) cells and LXR beta-UAS-*bla* HEK 293T cells contain a human Liver X receptor beta ligand-binding domain/Gal4 DNA binding domain chimera stably integrated into the CellSensor® UAS-*bla* HEK 293T cell line. CellSensor® UAS-*bla* HEK 293T contains a beta-lactamase reporter gene under control of a UAS response element stably integrated into HEK 293T cells. LXR beta HEK 293T DA cells and LXR beta-UAS-*bla* HEK 293T cells have been functionally validated for Z'-Factor and EC<sub>50</sub> concentrations of TO90131.



Dose response of LXR beta HEK 293T DA cells and LXR beta-UAS-*bla* HEK 293T cells to TO901317.

## 2. Overview of GeneBLAzer® Beta-Lactamase Reporter Technology

GeneBLAzer® Beta-lactamase Reporter Technology provides a highly accurate, sensitive, and easy-to-use method of monitoring cellular response to drug candidates or other stimuli (1). The core of the GeneBLAzer® Technology is a Fluorescence Resonance Energy Transfer (FRET) substrate that generates a ratiometric reporter response with minimal experimental noise. In addition to the dual-color (blue/green) readout of stimulated and unstimulated cells, this ratiometric method reduces the absolute and relative errors that can mask the underlying biological response of interest. Such errors include variations in cell number, transfection efficiency, substrate concentration, excitation path length, fluorescence detectors, and volume changes. The GeneBLAzer® Beta-lactamase Reporter Technology has been proven in high-throughput screening (HTS) campaigns for a range of target classes, including G-protein coupled receptors (GPCRs) (2, 3), nuclear receptors (4-6) and kinase signaling pathways (7).

## 3. Materials Supplied

| Product:  | Name  | Size    | Catalog # |
|---|---|---------|-----------|
|   | <b>GeneBLAzer® LXR beta HEK 293T DA Assay Kit</b><br>Each system contains sufficient division-arrested cells and substrate to assay one 384-well plate. Includes: <ul style="list-style-type: none"> <li>• LXR beta HEK 293T DA cells (K1415A)</li> <li>• LiveBLAzer™-FRET B/G Loading Kit, 70 µg</li> <li>• Protocol</li> <li>• Certificate of Analysis</li> </ul> | 1 plate | K1415     |
|   | <b>GeneBLAzer® LXR beta-UAS-<i>bla</i> HEK 293T cells</b><br>Includes: <ul style="list-style-type: none"> <li>• LXR beta-UAS-<i>bla</i> HEK 293T cells (K1131)</li> <li>• Protocol</li> <li>• Certificate of Analysis</li> </ul>  | 1 tube  | K1699     |
| <b>Shipping Condition:</b>                                  | Dry ice   |         |           |
| <b>Storage Condition of Cells:</b>                          | <b>Short Term:</b> -80°C<br><b>Long Term:</b> -80°C for at least 1 day followed by Liquid Nitrogen  |         |           |
| <b>Growth Properties of Non-Division-arrested Cells:</b>    | Adherent  |         |           |
| <b>Cell Phenotype:</b>                                      | Epithelial  |         |           |
| <b>Selection Marker(s) for Non-Division arrested cells:</b> | Blasticidin (15 µg/mL), Hygromycin (80 µg/mL)<br><b>Note:</b> HEK 293T cells contain the large T antigen and are thus Geneticin® resistant.   |         |           |
| <b>Mycoplasma Testing:</b>                                  | Negative  |         |           |
| <b>BioSafety Level:</b>                                     | 2   |         |           |

## 4. Materials Required

Use the table below to determine the additional media and reagents required for use with each kit:

| Media/Reagents   | Recommended Source | Part #  | Required Separately?                   |   |
|--|--------------------|---|--|---|
|  |                    |   | LXR beta HEK 293T DA Assay Kit (K1415) | LXR beta-UAS- <i>bla</i> HEK 293T cells (K1699) |
| LiveBLAzer <sup>™</sup> -FRET B/G Loading Kit:<br>LiveBLAzer <sup>™</sup> -FRET B/G Substrate (CCF4-AM)<br>DMSO for Solution A<br>Solution B<br>Solution C | Invitrogen         | K1427 (70 µg)<br>K1095 (200 µg)<br>K1096 (1 mg)<br>K1030 (5 mg) | No (included in kit)                   | Yes   |
| Recovery <sup>™</sup> Cell Culture Freezing Medium   | Invitrogen         | 12648-010   | No                                     | Yes   |
| DMEM (high-glucose), with GlutaMAX <sup>™</sup>  | Invitrogen         | 10569-010   | No                                     | Yes   |
| DMSO   | Fluka              | 41647   | Yes                                    | Yes   |
| Phenol red-free DMEM   | Invitrogen         | 21063-029   | Yes                                    | Yes   |
| Fetal bovine serum (FBS), charcoal-stripped  | Invitrogen         | 12676-011   | Yes                                    | Yes   |
| Fetal bovine serum (FBS), dialyzed, tissue-culture grade ( <b>DO NOT SUBSTITUTE!</b> )   | Invitrogen         | 26400-036   | No                                     | Yes   |
| Non-essential amino acids (NEAA)   | Invitrogen         | 11140-050   | Yes                                    | Yes   |
| Sodium Pyruvate  | Invitrogen         | 11360-070   | Yes                                    | Yes   |
| Penicillin/Streptomycin (antibiotics)  | Invitrogen         | 15140-122   | Yes                                    | Yes   |
| Phosphate-buffered saline without calcium and magnesium [PBS(-)]   | Invitrogen         | 14190-136   | No                                     | Yes   |
| HEPES (1 M, pH 7.3)  | Invitrogen         | 15630-080   | No                                     | Yes   |
| TO901317   | Cayman Chemical    | 71810   | Yes                                    | Yes   |
| 0.05% Trypsin/EDTA   | Invitrogen         | 25300-054   | No                                     | Yes   |
| Hygromycin (antibiotic)  | Invitrogen         | 10687-010   | No                                     | Yes   |
| Blasticidin (antibiotic)   | Invitrogen         | R210-01   | No                                     | Yes   |
| Matrigel <sup>™</sup> matrix, growth factor reduced  | BD                 | 356230  | No                                     | Yes   |

The following tables list materials required for use with all kits:

| Consumables  | Recommended Source | Part # |
|--|--------------------|--------|
| Black-wall, clear-bottom, 384-well assay plates (with low fluorescence background) | Corning            | 3712   |
| Compressed air   | Various            | ---    |

| Equipment   | Recommended Source  |
|---|---------------------|
| Fluorescence plate reader with bottom-read capabilities   | Various             |
| Filters if required for plate reader (see <b>Section 7.4.1</b> )                                | Chroma Technologies |
| <b>Optional:</b> Epifluorescence- or fluorescence-equipped microscope, with appropriate filters | Various             |
| <b>Optional:</b> Microplate centrifuge  | Various             |

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## 5. Detailed Cell Handling Procedures

**Note:** Division-arrested (DA) cells have different thawing procedures than non-DA cells. Refer to the instructions below for your particular application.

**Note:** Refer to **Section 6, Media Requirements** for specific media recipes.

### 5.1 DA Cells Thawing Method

**Note:** Once cells are thawed per the instructions below, cells must be counted and the density adjusted to the appropriate level as specified in **Section 7, Assay Procedure**, prior to analysis.

1. Rapidly thaw the vial of cells by placing at 37°C in a water bath with gentle agitation for 1–2 minutes. Do not submerge vial in water.
2. Decontaminate the vial by wiping with 70% ethanol before opening in a Class II biological safety cabinet.
3. Transfer the vial contents drop-wise into 10 mL of Assay Medium in a sterile 15-mL conical tube.
4. Centrifuge cells at 200 × g for 5 minutes.
5. Aspirate supernatant and resuspend the cell pellet in 1 mL fresh Assay Medium.
6. Count the cells.
7. Proceed to **Section 7, Assay Procedure**. Adjust the cell density with Assay Medium to the appropriate cell density as specified in Section 7.

### 5.2 Non-DA Cells

#### 5.2.1 Thawing Method

**Note:** Cells are shipped to you on dry ice and as such may require a short period of time prior to full recovery and normal growth.

1. Precoat a T25 flask with 1 mL 1X Matrigel<sup>™</sup> matrix. Incubate 15 min. at 37°C. Aspirate.
2. Place 9 mL of Thawing Medium into the precoated T25 flask. Place the flask in a humidified 37°C/5% CO<sub>2</sub> incubator for 15 minutes to allow medium to equilibrate to the proper pH and temperature.
3. Remove the vial of cells to be thawed from liquid nitrogen and rapidly thaw by placing at 37°C in a water bath with gentle agitation for 1–2 minutes. Do not submerge vial in water.
4. Decontaminate the vial by wiping with 70% ethanol before opening in a Class II biological safety cabinet.
5. Transfer the vial contents drop-wise into 10 mL of Thawing Medium in a sterile 15-mL conical tube.
6. Centrifuge cells at 200 × g for 5 minutes.
7. Aspirate supernatant and resuspend the cell pellet in 1 mL of fresh Thawing Medium.
8. Count Cells.
9. Transfer ~1 × 10<sup>6</sup> cells to the T25 tissue culture flask (~40,000 cells/cm<sup>2</sup>) containing pre-equilibrated Thawing Medium and place flask in the humidified 37°C/5% CO<sub>2</sub> incubator.
10. At first passage, switch to Growth Medium.

#### 5.2.2 Propagation Method

1. Passage or feed cells at least twice a week. Maintain cells between 5% and 95% confluence. **Do not allow cells to reach confluence.**
2. All flasks must be coated with 1X Matrigel<sup>™</sup> matrix (1 mL for a T25 flask, 3 mL for a T75 flask, 5 mL for a T175 flask, and 7 mL for T225 flask) and incubated 15 min. in a humidified 37°C/5% CO<sub>2</sub> incubator before plating. After coating and incubation, aspirate the Matrigel<sup>™</sup> matrix and plate cells.
3. To passage cells, aspirate medium, rinse once in PBS, add Trypsin/EDTA (1 mL for a T25 flask, 3 mL for a T75 flask, 5 mL for a T175 flask, and 7 mL for T225 flask) and swirl to coat the cells evenly. Cells usually detach after ~2–5 minutes exposure to Trypsin/EDTA. Add an equal volume of Growth Medium to inactivate Trypsin.
4. Verify under a microscope that cells have detached and clumps have completely dispersed.
5. Centrifuge cells at 200 × g for 5 minutes and resuspend in Growth Medium.

#### 5.2.3 Freezing Method

1. Harvest the cells as described in **Subsection 5.2.2** (above), Step 2. After detachment, count the cells, centrifuge cells at 200 × g for 5 minutes, and resuspend in 4°C Freeze Medium to a density of 2 × 10<sup>6</sup> cells/mL.

2. Dispense 1.0-mL aliquots into cryogenic vials.
3. Place in an insulated container for slow cooling and store overnight at -80°C.
4. Transfer to liquid nitrogen the next day for storage.

## 6. Media Requirements

**Note:** Unless otherwise stated, have all media and solutions at least at room temperature (we recommend 37°C for optimal performance) before adding to cells. (Matrigel should be kept cold prior to coating flasks.)

**Note:** Make **NO MEDIA SUBSTITUTIONS**, as these cell lines have been specifically validated for optimal assay performance with these media. For non-DA cells, we recommend that you create and store an aliquot for back up.

**Note:** All media components can be added directly to the 500 mL bottle of base media (DMEM).

| Component                                    | Assay Medium<br>(DA and Non-DA cells) | Growth Medium<br>(Non-DA cells only) | Thawing Medium<br>(Non-DA cells only) | Freeze Medium<br>(Non-DA cells only) | 1X Matrigel™ Matrix<br>(Non-DA cells only) |
|--|---------------------------------------|--------------------------------------|---------------------------------------|--------------------------------------|--|
| DMEM with GlutaMAX™                          | --                                    | 90% (500 mL)                         | 90% (500 mL)                          | —                                    | —  |
| Phenol red-free DMEM                         | 98% (500 mL)                          | —                                    | —                                     | —                                    | 99.75%                                     |
| Dialyzed FBS (Do not substitute!)            | —                                     | 10% (50 ml)                          | 10% (50 mL)                           | —                                    | —  |
| Charcoal-stripped FBS                        | 2% (10 mL)                            | —                                    | —                                     | —                                    | —  |
| Sodium Pyruvate                              | 1 mM (5 mL)                           | —                                    | —                                     | —                                    | —  |
| NEAA   | 0.1 mM (5 mL)                         | 0.1 mM (5 mL)                        | 0.1 mM (5 mL)                         | —                                    | —  |
| HEPES (pH 7.3)                               | —                                     | 25 mM (12.5 mL)                      | 25 mM (12.5 mL)                       | —                                    | —  |
| Penicillin/<br>Streptomycin<br>(antibiotics) | 100 U/mL and<br>100 µg/mL<br>(5 mL)   | 100 U/mL and<br>100 µg/mL<br>(5 mL)  | 100 U/mL and<br>100 µg/mL<br>(5 mL)   | —                                    | —  |
| Hygromycin<br>(antibiotic)                   | —                                     | 80 µg/mL                             | —                                     | —                                    | —  |
| Blasticidin<br>(antibiotic)                  | —                                     | 15 µg/mL                             | —                                     | —                                    | —  |
| Recovery™ Cell Culture Freezing Medium       | —                                     | —                                    | —                                     | 100%                                 | —  |
| Matrigel™ matrix                             | —                                     | —                                    | —                                     | —                                    | 0.25%                                      |

## 7. Assay Procedure

The following instructions outline the recommended procedure for determining activity of compounds as modulators of LXR beta using LiveBLAzer<sup>™</sup>-FRET B/G Substrate as the readout. If alternative substrates are used (*e.g.*, ToxBLAzer<sup>™</sup> DualScreen or LyticBLAzer<sup>™</sup> Loading kits), follow the loading protocol provided with the product.

### 7.1 Quick Assay Reference Guides

For a more detailed assay protocol, see [Section 7.2](#).

#### Agonist Assay Quick Reference Guide

|  | Unstimulated Wells   | Stimulated Wells                                     | Cell-free Wells                       | Test Compound Wells                                  |
|--|--|--|---------------------------------------|--|
| <b>Step 1</b><br>Plate cells, incubate           | 32 $\mu$ L cells in Assay Medium (10,000 cells/well)   | 32 $\mu$ L cells in Assay Medium (10,000 cells/well) | 32 $\mu$ L Assay Medium (no cells)    | 32 $\mu$ L cells in Assay Medium (10,000 cells/well) |
| <b>Step 2</b><br>Add Agonist or Test Compounds   | 8 $\mu$ L Assay Medium with 0.5% DMSO  | 8 $\mu$ L 5X TO901317 in Assay Medium with 0.5% DMSO | 8 $\mu$ L Assay Medium with 0.5% DMSO | 8 $\mu$ L 5X Test Compounds in 0.5% DMSO             |
| <b>Step 3</b><br>Incubate cells                  | Incubate in a humidified 37°C/5% CO <sub>2</sub> incubator for 20 hours  |  |                                       |  |
| <b>Step 4</b><br>Prepare 6X Substrate Mix        | 6 $\mu$ L of 1 mM LiveBLAzer <sup>™</sup> -FRET B/G (CCF4-AM) Substrate + 60 $\mu$ L of solution B, mix. Add 934 $\mu$ L of Solution C, mix. |  |                                       |  |
| <b>Step 5</b><br>Add Substrate Mixture           | 8 $\mu$ L per well   |  |                                       |  |
| <b>Step 6</b><br>Incubate Substrate Mix. + cells | 2 hours at room temperature in the dark  |  |                                       |  |
| <b>Step 7</b><br>Detect activity                 | See <a href="#">Section 7.4</a>  |  |                                       |  |
| <b>Step 8</b><br>Analyze data                    | See <a href="#">Section 8</a>  |  |                                       |  |

#### Antagonist Assay Quick Reference Guide

|   | Unstimulated Wells   | Stimulated Wells                                      | Antagonist Control Wells                                | Cell-free Wells                       | Test Compound Wells   |
|---|--|---|---|---------------------------------------|---|
| <b>Step 1</b><br>Plate cells, incubate            | 32 $\mu$ L cells in Assay Medium (10,000 cells/well)   | 32 $\mu$ L cells in Assay Medium (10,000 cells/well)  | 32 $\mu$ L cells in Assay Medium (10,000 cells/well)    | 32 $\mu$ L Assay Medium (no cells)    | 32 $\mu$ L cells in Assay Medium (10,000 cells/well)        |
| <b>Step 2</b><br>Add Antagonist or Test Compounds | 4 $\mu$ L Assay Medium with 0.5% DMSO  | 4 $\mu$ L Assay Medium with 0.5% DMSO                 | 4 $\mu$ L 10X antagonist in Assay Medium with 0.5% DMSO | 4 $\mu$ L Assay Medium with 0.5% DMSO | 4 $\mu$ L 10X Test Compounds in Assay Medium with 0.5% DMSO |
| <b>Optional Step:</b>                             | Incubate plate with Antagonist for 30 minutes before proceeding  |   |   |                                       |   |
| <b>Step 3</b><br>Add Agonist                      | 4 $\mu$ L Assay Medium with 0.5% DMSO  | 4 $\mu$ L 10X TO901317 in Assay Medium with 0.5% DMSO | 4 $\mu$ L 10X TO901317 in Assay Medium with 0.5% DMSO   | 4 $\mu$ L Assay Medium with 0.5% DMSO | 4 $\mu$ L 10X TO901317 in Assay Medium with 0.5% DMSO       |
| <b>Step 4</b><br>Incubate cells                   | Incubate in a humidified 37°C/5% CO <sub>2</sub> incubator for 20 hours  |   |   |                                       |   |
| <b>Step 5</b><br>Prepare 6X Substrate Mix         | Add 6 $\mu$ L of 1 mM LiveBLAzer <sup>™</sup> -FRET B/G (CCF4-AM) substrate + 60 $\mu$ L of solution B, mix. Add 934 $\mu$ L of Solution C, mix. |   |   |                                       |   |
| <b>Step 6</b><br>Add Substrate Mixture            | 8 $\mu$ L per well   |   |   |                                       |   |
| <b>Step 7</b><br>Incubate Mixture                 | 2 hours at room temperature in the dark  |   |   |                                       |   |
| <b>Step 8</b><br>Detect activity                  | See <a href="#">Section 7.4</a>  |   |   |                                       |   |
| <b>Step 9</b><br>Analyze data                     | See <a href="#">Section 8</a>  |   |   |                                       |   |

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## 7.2 Detailed Assay Protocol

Plate layouts and experimental outlines will vary; in screening mode, we recommend using at least three wells for each control: Unstimulated Control, Stimulated Control, and Cell-free Control.

**Note:** Some solvents may affect assay performance. Assess the effects of solvent before screening. The cell stimulation procedure described below is carried out in the presence of 0.1% DMSO to simulate the effect that a Test Compound's solvent might have on the assay. If you use other solvents and/or solvent concentrations, optimize the following assay conditions appropriately.

### 7.2.1 Precautions

- Work on a dust-free, clean surface. Always handle the 384-well, black-wall, clear-bottom assay plate by the sides; do not touch the clear bottom of the assay plate.
- If pipetting manually, you may need to centrifuge the plate briefly at room temperature (for 1 minute at  $14 \times g$ ) after additions to ensure all assay components are on the bottom of the wells.

### 7.2.2 Plating Cells

#### DA Cells

1. Thaw DA cells into Assay Medium and count (as described in **Section 5.1**). Dilute cells to a density of  $3.1 \times 10^5$  cells/mL in Assay Medium.
2. Add 32  $\mu$ L per well of the Assay Medium to the Cell-free Control wells. Add 32  $\mu$ L per well (10,000 cells/well) of the cell suspension to the Test Compound wells, the Unstimulated Control wells, and Stimulated Control wells. Proceed to **Section 7.2.3** for an Agonist assay or **Section 7.2.4** for an Antagonist assay.

#### Non-DA Cells

1. Harvest non-DA cells from culture at 50–75% confluency. Spin down cells and suspend cells in Assay Medium and count. Dilute cells to a density of  $3.1 \times 10^5$  cells/mL in Assay Medium.
2. Add 32  $\mu$ L per well of the Assay Medium to the Cell-free Control wells. Add 32  $\mu$ L per well (10,000 cells/well) of the cell suspension to the Test Compound wells, the Unstimulated Control wells, and Stimulated Control wells. Proceed to **Section 7.2.3** for an Agonist assay or **Section 7.2.4** for an Antagonist assay.

### 7.2.3 Agonist Assay Plate Setup

**Note:** This subsection provides directions for performing an Agonist assay. See **Section 7.2.4** for directions for performing an Antagonist assay.

1. Prepare a stock solution of 0.5% DMSO in Assay Medium.
2. Prepare a 5X stock of Test Compounds in Assay Medium with 0.5% DMSO (or if test compound is dissolved in DMSO, make sure the DMSO concentration for the 5X solution is 0.5%)
3. Prepare a 5X stock of TO901317 in Assay Medium with 0.5% DMSO. We recommend running a dose response curve to determine the optimal concentration for the TO901317 solution.
4. Add 8  $\mu$ L of the stock solution of 0.5% DMSO in Assay Medium to the Unstimulated Control and Cell-free Control wells.
5. Add 8  $\mu$ L of the 5X stock solution of TO901317 to the Stimulated Control wells.
6. Add 8  $\mu$ L of the 5X stock of Test Compounds to the Test Compound wells.
7. Incubate the Agonist assay plate in a humidified 37°C/5% CO<sub>2</sub> incubator for 20 hours. Then proceed to **Section 7.3** for Substrate Loading and Incubation.

### 7.2.4 Antagonist Assay Plate Setup

**Note:** This subsection provides directions for performing an Antagonist assay. See **Section 7.2.3** for directions for performing an Agonist assay.

1. Prepare a stock solution of 0.5% DMSO in Assay Medium.
2. Prepare a 10X stock of Test Compounds in Assay Medium with 0.5% DMSO (or if test compound is dissolved in DMSO, make sure the DMSO concentration for the 5X solution is 0.5%).
3. Prepare a 10X stock of TO901317 in Assay Medium with 0.5% DMSO. We recommend running a dose response curve to determine the optimal agonist concentration. For antagonist assays, we recommend stimulating cells with an agonist concentration in the EC<sub>50</sub>–EC<sub>80</sub> range.
4. Prepare a 10X stock of antagonist in Assay Medium with 0.5% DMSO. We recommend running a dose response curve to determine the optimal inhibition concentration for the Antagonist solution.
5. Add 4  $\mu$ L of the 10X stock of Test Compounds to the Test Compound wells.

6. Add 4  $\mu$ L of the stock solution of 0.5% DMSO in Assay Medium to the Stimulated Control wells, the Unstimulated Control wells, and the Cell-free Control wells.
7. Add 4  $\mu$ L of the 10X stock of antagonist in Assay Medium with 0.5% DMSO to the Antagonist Control wells.
8. If desired, incubate the Test Compounds with the cells in a humidified 37°C/5% CO<sub>2</sub> incubator before proceeding. Typically, a 30-minute incubation is sufficient.
9. Add 4  $\mu$ L of the 10X stock solution of TO901317 to the Test Compound wells, the Stimulated Control wells, and the Antagonist Control wells.
10. Add 4  $\mu$ L of Assay Medium with 0.5% DMSO to the Unstimulated Control and Cell-free Control wells.
11. Incubate the Antagonist assay plate in a humidified 37°C/5% CO<sub>2</sub> incubator for 20 hours. Then proceed to **Section 7.3** for Substrate Loading and Incubation.

### 7.3 Substrate Preparation, Loading and Incubation

This protocol is designed for loading cells with LiveBLAzer<sup>™</sup>-FRET B/G Substrate Mixture (CCF4-AM) Substrate Mixture. If you use alternative substrates, follow the loading protocol provided with the substrate.

Prepare LiveBLAzer<sup>™</sup>-FRET B/G Substrate Mixture (CCF4-AM) Substrate Mixture and load cells in the absence of direct strong lighting. Turn off the light in the hood.

1. Prepare Solution A: 1 mM LiveBLAzer<sup>™</sup>-FRET B/G Substrate (CCF4-AM) Substrate Mixture in dry DMSO by adding 912  $\mu$ L of DMSO per mg of dry substrate. Store the aliquots of the stock solution at -20°C until use. The molecular weight of the LiveBLAzer<sup>™</sup>-FRET B/G Substrate (CCF4-AM) is 1096 g/mol.
2. Prepare 6X Loading Solution:
  - a. Add 6  $\mu$ L of Solution A to 60  $\mu$ L of Solution B and vortex.
  - b. Add 934  $\mu$ L of Solution C to the above solution and vortex.

**Note:** If more than 1 mL 6X Substrate Mixture is needed, scale up the amount of each solution proportionally

3. Remove assay plate from the humidified 37°C/5% CO<sub>2</sub> incubator.
 

**Note:** Handle the plate gently and do not touch the bottom.
4. Add 8  $\mu$ L of the 6X Substrate Mixture to each well.
5. Cover the plate to protect it from light and evaporation.
6. Incubate at room temperature for 2 hours.

### 7.4 Detection

Make measurements at room temperature from the bottom of the wells, preferably in 384-well, black-wall, clear-bottom assay plates with low fluorescence background. Before reading the plate, remove dust from the bottom with compressed air.

#### 7.4.1 Instrumentation, Filters, and Plates

- Fluorescence plate reader with bottom reading capabilities.
- Recommended filters for fluorescence plate reader:
  - Excitation filter: 409/20 nm
  - Emission filter: 460/40 nm
  - Emission filter: 530/30 nm

#### 7.4.2 Reading an Assay Plate

1. Set the fluorescence plate reader to bottom-read mode with optimal gain and 5 reads.
2. Allow the lamp in the fluorescence plate reader to warm up for at least 10 minutes before making measurements.
3. Use the following filter selections:

|                           | Scan 1                                   | Scan 2                                   |
|---------------------------|--|--|
| <b>Purpose:</b>           | Measure fluorescence in the Blue channel | Measure FRET signal in the Green channel |
| <b>Excitation filter:</b> | 409/20 nm                                | 409/20 nm                                |
| <b>Emission filter:</b>   | 460/40 nm                                | 530/30 nm                                |



## 8. Data Analysis

---

### 8.1 Background Subtraction and Ratio Calculation

We recommend that you subtract the background for both emission channels (460 nm and 530 nm).

1. Use the assay plate layout to identify the location of the Cell-free Control wells. These Control wells are used for background subtraction.
2. Determine the average emission from the Cell-free Control wells at both 460 nm (Average Blue Background) and 530 nm (Average Green Background).
3. Subtract the Average Blue background from all of the Blue emission data.
4. Subtract the Average Green background from all of the Green emission data.
5. Calculate the Blue/Green Emission Ratio for each well, by dividing the background-subtracted Blue emission values by the background-subtracted Green emission values.

**Note:** You may also calculate response ratio to know your assay window. The response ratio is calculated as the Blue/Green Emission Ratio of the TO901317 –Stimulated wells divided by the Blue/Green Emission Ratio of the unstimulated wells. Generally, a response ratio of >3 has been shown to yield a  $Z' \geq 0.6$ .

### 8.2 Visual Observation of Intracellular Beta-lactamase Activity Using LiveBLAzer<sup>™</sup>-FRET B/G Substrate (CCF4-AM)

**Note:** Microscopic visualization of cells will cause photobleaching. Always read the assay plate in the fluorescence plate reader before performing microscopic visualization.

An inverted microscope equipped for epifluorescence and with either a xenon or mercury excitation lamp may be used to view the LiveBLAzer<sup>™</sup>-FRET B/G Substrate (CCF4-AM) signal in cells. To visually inspect the cells, you will need a long-pass filter passing blue and green fluorescence light, so that your eye can visually identify whether the cells are fluorescing green or blue.

Recommended filter sets for observing beta-lactamase activity are described below and are available from Chroma Technologies (800-824-7662, [www.chroma.com](http://www.chroma.com)).

#### Chroma Set # 41031

|                    |                         |
|--------------------|-------------------------|
| Excitation filter: | HQ405/20x (405 ± 10)    |
| Dichroic mirror:   | 425 DCXR                |
| Emission filter:   | HQ435LP (435 long-pass) |

Filter sizes vary for specific microscopes and need to be specified when the filters are ordered. For epifluorescence microscopes, a long-pass dichroic mirror is needed to separate excitation and emission light and should be matched to the excitation filter (to maximally block the excitation light around 405 nm, yet allow good transmission of the emitted light).

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## 10. Purchaser Notification

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