

Membrane Permeability/Dead Cell Apoptosis Kit with PO-PRO®-1 and 7-aminoactinomycin D for Flow Cytometry

Catalog no. V35123

Table 1. Contents and storage information.

Material	Amount	Composition	Storage*	Stability
PO-PRO®-1 dye (Component A)	500 µL	1 mM solution in DMSO	<ul style="list-style-type: none"> • 2–6°C • Dessicate • Protect from light 	When stored as directed this kit is stable for 6 months.
7-aminoactinomycin D (7-AAD, Component B)	200 µL	1 mg/mL solution in DMSO		
*For long-term storage, store the vial of PO-PRO®-1 dye at ≤-20°C. The PO-PRO®-1 dye and 7-AAD are light sensitive and may be handled in normal room light, but avoid prolonged exposure to light.				
Number of assays: Sufficient material is supplied for 200 flow cytometry assays each having 2×10^5 to 1×10^6 cells in a 1 mL volume.				
Approximate fluorescence excitation/emission maxima: PO-PRO®-1: 434/456 in nm; 7-AAD: 546/647 in nm.				

Introduction

Apoptosis is a carefully regulated process of cell death that occurs as a normal part of development. Inappropriately regulated apoptosis is implicated in disease states, such as Alzheimer's disease and cancer. Apoptosis is distinguished from necrosis, or accidental cell death, by characteristic morphological and biochemical changes, including compaction and fragmentation of the nuclear chromatin, shrinkage of the cytoplasm and loss of membrane asymmetry.^{1–5} Furthermore, during apoptosis the cytoplasmic membrane becomes slightly permeant. Certain dyes, such as the violet fluorescent PO-PRO™-1 dye can enter apoptotic cells, whereas other dyes, such as the red fluorescent dye, 7-aminoactinomycin D (7-AAD), cannot. The use of YO-PRO®-1 dye (closely related to PO-PRO™-1 dye) with propidium iodide provides a sensitive indicator for apoptosis.^{6–9} PO-PRO™-1 dye and 7-AAD provide analogous performance. In addition, annexin V gives poor results with trypsinized cells while PO-PRO™-1 dye provides the same efficiency with trypsinized cells as it does with suspension cells.

The Membrane Permeability/Dead Cell Apoptosis Kit with PO-PRO®-1 and 7-aminoactinomycin D for flow cytometry provides a rapid and convenient assay for apoptosis. The kit contains ready-to-use solutions of both PO-PRO®-1 and 7-AAD dyes. After staining a cell population with PO-PRO®-1 dye and 7-AAD, apoptotic cells show violet fluorescence, dead cells show violet and red fluorescence, and live cells show little or no fluorescence (Figure 1). These populations can easily be distinguished by a flow cytometer that uses both a violet laser and the 488 nm line of an argon-ion laser for excitation.

We have optimized this assay using Jurkat cells, a human T-cell leukemia line, treated with camptothecin to induce apoptosis. Some modifications may be required for use with other cell types. Because no single parameter defines apoptosis in all systems, we strongly suggest using a combination of different measurements for reliable detection of apoptosis. Refer to

Before Starting

Materials Required but Not Provided

- Samples (appropriate sample concentrations range from 2×10^5 to 1×10^6 cells/mL)
- Inducing agent
- 2 mM hydrogen peroxide
- Phosphate buffered saline (PBS)
- Deionized water

Caution

No data are available addressing the mutagenicity or toxicity of 7-AAD or PO-PRO[®]-1 dye. Because these reagents bind to nucleic acids, treat them as potential mutagens and handle with appropriate care. Handle the DMSO stock solutions with particular caution as DMSO is known to facilitate the entry of organic molecules into tissues.

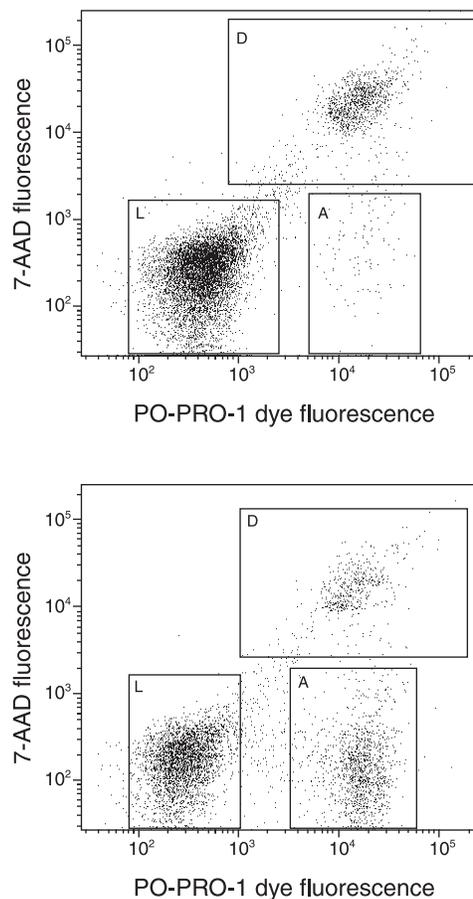


Figure 1. Jurkat cells (human T-cell leukemia) treated with 10 μ M camptothecin for four hours (bottom panel) or untreated (as control, top panel). Cells were then treated with the reagents in the kit and analyzed by flow cytometry using 405 nm and 488 nm excitation. Note that the camptothecin-treated cells have a higher percentage of apoptotic cells (indicated by an 'A') than the basal level of apoptosis seen in the control cells. L = live cells, D = dead cells.

Experimental Protocol

We have optimized this assay using Jurkat cells treated with camptothecin to induce apoptosis. Some modifications may be required for use with other cell types.

1. Induce apoptosis in cells using the desired method. Prepare a negative control by incubating cells in the absence of inducing agent.
2. Harvest the cells after the incubation period, wash in cold phosphate-buffered saline (PBS), and adjust the cell density to $\sim 1 \times 10^6$ cells/mL in PBS. For each assay, use a 1 mL volume.
3. Add 2.5 μ L PO-PRO[®]-1 stock solution (Component A) and 1 μ L 7-AAD stock solution (Component B) to each 1 mL of cell suspension.
4. Incubate the cells on ice for 30 minutes.
5. As soon as possible after the incubation period, analyze the stained cells by flow cytometry, using violet and 488 nm excitation and measuring the fluorescence emission using 440 nm and 670 nm bandpass filters (or their near equivalents). The population should separate into three groups: live cells will show only a low level of fluorescence, apoptotic cells will show violet fluorescence and necrotic cells will show both red and violet fluorescence (see Figure 1). Confirm the flow cytometry results by viewing the cells under a fluorescence microscope using the appropriate filters.

References

1. Immunol Cell Biol 76, 1 (1998); 2. Cytometry 27, 1 (1997); 3. J Pharmacol Toxicol Methods 37, 215 (1997); 4. FASEB J 9, 1277 (1995); 5. Am J Pathol 146, 3 (1995); 6. Cancer Res 57, 3804 (1997); 7. Blood 87, 4959 (1996); 8. J Exp Med 182, 1759 (1995); 9. J Immunol Methods 185, 249 (1995).

Product List

Current prices may be obtained from our website or from our Customer Service Department.

Cat. no.	Product Name	Unit Size
V35123	Membrane Permeability/Dead Cell Apoptosis Kit with PO-PRO [®] -1 and 7-aminoactinomycin D *for flow cytometry* *200 assays* . . .	1 kit

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